

## Amphidinolide U, Novel 20-Membered Macrolide from Marine Dinoflagellate *Amphidinium* sp.

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Received 30 September 1999; accepted 15 October 1999

Abstract; A novel 20-membered macrolide, amphidinolide U (1), has been isolated from a marine dinoflagellate Amphidinium sp. and the structure was elucidated on the basis of spectroscopic data. The relative stereochemistry of C-15, C-18, and C-19 was deduced from NOESY correlations, while the absolute configurations at C-8 and C-24 were assigned as both S on the basis of modified Mosher's method. © 1999 Elsevier Science Ltd. All rights reserved.

keywords; marine dinoflagellate, macrolide, tetrahydrofuran

Amphidinolides A ~ H and J ~ S are a series of unique macrolides obtained from marine dinoflagellates of the genus Amphidinium, which are symbionts of Okinawan marine accel flatworms Amphiscolops spp. Our continuing search for bioactive secondary metabolites from laboratory-cultured marine dinoflagellates<sup>2-5</sup> resulted in the isolation of a novel 20-membered macrolide, amphidinolide U (1), from extracts of another strain (Y-56) of the dinoflagellate Amphidinium sp. Here we describe the isolation and structure elucidation of 1.

The acoel flatworm *Amphiscolops* sp. was collected off Zanpa, Okinawa, from which an associated dinoflagellate *Amphidinium* sp. was isolated and mass cultured unialgally at 25 °C for 2 weeks in a seawater medium enriched with 1% ES supplement. The harvested algal cells (420 g, wet weight, from 580 L of culture) were extracted with MeOH/toluene (3:1), and the extracts were partitioned between toluene and water. The toluene-soluble materials were subjected to silica gel (CHCl<sub>3</sub>/MeOH) and C<sub>18</sub> columns (CH<sub>3</sub>CN/H<sub>2</sub>O) followed by C<sub>18</sub> HPLC (CH<sub>3</sub>CN/H<sub>2</sub>O) to afford

Stereochemistries of C-15, C-18, and C-19 are relative.

positn.	$\delta_{\rm c}$		HMBC (H)				
1	170.8	S					2, 19
2	42.8	t	$3.07^{a}$	m			30
3	142.7	S					2, 30
4	35.3	t	$2.13^{a}$	m			2, 5, 30
4 5	26.4	t	2.20	m	2.17	m	4, 6
6 7	125.6	d	5.45	m			4, 31
7	135.4	S					$9b^c, 31$
8	73.1	d	4.33	m			9b°, 31
8-OH			3.17	br			
9	44.3	t	2.96	dd, 4.6, 12.7	2.72	dd, 6.5, 12.7	11
10	213.4	S					9, 32
11	41.7	d	3.05	m			$12a^{c}$ , 32
12	46.1	t	2.98	dd, 8.4, 18.1	2.36	dd, 4.3, 18.1	32
13	206.8	S					12, 14
14	48.5	t	2.70	dd, 8.7, 15.8	2.44	dd, 3.6, 15.8	16b <sup>c</sup>
15	75.2	d	4.31	m			14a <sup>c</sup> , 16b, 18
16	32.2	t	2.10	m	1.49	m	14 <b>a</b> °
17	28.2	t	1.96	m	1.64	m	.=
18	79.77	d	4.09	brq, 7.0			17b°, 19
19	77.0	d	5.22	brt, 7.3			17b°, 20, 21
20	127.8	d	5.60	dd, 7.9, 15.1			19, 22
21	130.6	d	6.54	dd, 11.0, 15.1			19, 22
22	125.2	d	6.13	brd, 11.0			20, 24, 33
23	139.9	S					24, 33
$24^d$	79.79	d	4.48	S			22, 26, 33, 34
25	149.2	S					24, 26, 27, 34a <sup>c</sup>
26	31.5	t	1.93	m	1.86	m	27, 34
27	30.1	t	1.41"	m			26, 29
28	22.5	t	1.31"	m			27, 29
29	14.0	q	$0.89^{b}$	t, 7.3			27
30	113.7	ť	4.92"	S			2
31	13.0	q	1.67 <sup>b</sup>	S			
32	16.7	q	1.09"	d, 7.1			12a <sup>c</sup>
33	12.6	q	1.66 <sup>b</sup>	S			22, 24
34	110.3	ť	5.12	S	4.95	S	24, 26

Table 1. <sup>1</sup>H and <sup>13</sup>C NMR Data of Amphidinolide U (1) in CDCl<sub>3</sub>.

 $^a$ 2H.  $^b$ 3H.  $^c$ a and b denote low-field and high-field resonances respectively of a geminal pair.  $^d$ 24-OH was not detected.

amphidinolide U (1, 0.8 mg, 0.0002 %, wet weight) together with two known macrolides, amphidinolides  $C^6$  (3, 0.0009 %) and  $F^7$  (0.0006 %).

Amphidinolide U (1) had the molecular formula of  $C_{34}H_{50}O_7$  as revealed by HRFABMS [m/z 593.3462, (M+Na)<sup>+</sup>, +1.7 mmu]. IR absorptions at 3400 and 1710 cm<sup>-1</sup> indicated the presence of hydroxyl(s) and carbonyl group(s), respectively. The UV spectrum showed the absorption at 230 nm ( $\epsilon$  20000) due to a conjugated diene chromophore. <sup>1</sup>H and <sup>13</sup>C NMR data (Table 1) disclosed the existence of two ketones, an ester carbonyl, four sp<sup>2</sup> quaternary carbons, four sp<sup>2</sup> methines, two sp<sup>2</sup> methylenes, six sp<sup>3</sup> methines (five of them bearing an oxygen atom), eleven sp<sup>3</sup> methylenes, and four methyls (two of them attached to olefins). Thus accounting for eight out of ten unsaturations, amphidinolide U (1) was inferred to contain two rings. Interpretation of the <sup>1</sup>H-<sup>1</sup>H COSY and TOCSY spectra revealed proton connectivities of the following partial structures: (a) from H<sub>2</sub>-2 to H<sub>2</sub>-30, (b) from H<sub>2</sub>-4 to H<sub>3</sub>-31, (c) from 8-OH to H<sub>2</sub>-9, (d) from H<sub>3</sub>-32 to H<sub>2</sub>-12, (e) from H<sub>2</sub>-14 to H<sub>3</sub>-

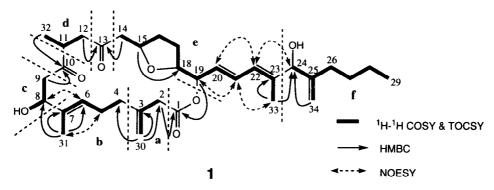


Figure 1. Selected 2D NMR Correlations for Amphidinolide U (1)

33, and (f) from H-24 to  $H_3$ -29 and  $H_2$ -34 (Figure 1). Connections among partial structures **a**-f and three quaternary carbons (C-1, C-10, and C-13) were assigned on the basis of  ${}^{1}H_{-}^{13}C$  long-range correlations observed in the HMBC spectrum. HMBC correlations from  $H_2$ -30 to C-2 ( $\delta_{\rm C}$  42.8), C-3 ( $\delta_{\rm C}$  142.7), and C-4 ( $\delta_{\rm C}$  35.3) suggested the connectivity between partial structures **a** and **b** through an *exo*-methylene at C-3. Connections among units **b**, **c**, **d**, **e**, and **f** were deduced from the following HMBC correlations;  $H_3$ -31/C-6,  $H_3$ -31/C-7,  $H_3$ -31/C-8,  $H_2$ -9/C-10,  $H_3$ -32/C-10,  $H_2$ -12/C-13,  $H_2$ -14/C-13,  $H_3$ -33/C-23, and  $H_3$ -33/C-24. The existence of an ester linkage between C-1 and C-19 was implied by HMBC correlations from  $H_2$ -2 and H-19 to C-1 ( $\delta_{\rm C}$  170.8). The HMBC cross-peak from H-15 to C-18 suggested that C-15 and C-18 were linked through an oxygen to form a tetrahydrofuran ring. Geometries of three internal olefins at C-6-C-7, C-20-C-21, and C-22-C-23 were assigned as all *E* on the basis of NOESY cross-peaks for  $H_2$ -5/ $H_3$ -31, H-6/H-8, H-19/H-21, H-20/H-22, H-21/ $H_3$ -33, and H-22/H-24. Thus the gross structure of amphidinolide U was elucidated to be **1**.

NOESY correlations were observed for H-14a/H-16a, H-14a/H-17a, H-15/H-17b, H-16a/H-18, H-17a/H-19, and H-17b/H-20, indicating that relative stereochemistries between H-15 and H-18 and between H-18 and H-19 were *anti*- and *syn*-oriented, respectively (Figure 2). The absolute

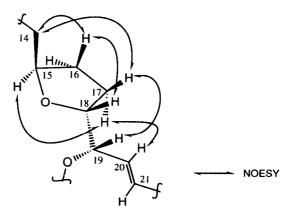


Figure 2. Relative Stereochemistry of Tetrahydrofuran Ring in Amphidinolide U (1)

Figure 3.  $\Delta\delta$  values [ $\Delta\delta$  (in ppm) =  $\delta_S$  - $\delta_R$ ] obtained for (*S*)- and (*R*)-MTPA ester (**2a** and **2b**, respectively) of amphidinolide U (**1**). Stereochemistries of C-15, C-18, and C-19 are relative.

configurations at C-8 and C-24 were assigned by modified Mosher's method<sup>8</sup> as follows (Figure 3). Amphidinolide U (1) was treated with (R)-(-)- or (S)-(+)-2-methoxy-2-trifluoromethyl-2-phenylacetyl chloride (MTPACl) to afford (S)-(-)- and (R)-(+)-MTPA ester (2a and 2b, respectively) of 1.  $\Delta\delta$  values of H-6 (+0.06) and H<sub>3</sub>-31 (+0.11) showed positive signs, while those of H<sub>2</sub>-9 (-0.10 and -0.06) H<sub>2</sub>-12 (-0.05 and -0.05), and H<sub>3</sub>-32 (-0.02) were negative values, thus indicating 8*S*-configuration. On the other hand, 24*S*-configuration was determined on the basis of  $\Delta\delta$  values of H-20 (+0.11), H-21 (+0.06), H-22 (+0.13), H<sub>2</sub>-26 (-0.07),H<sub>2</sub>-27 (-0.06), H<sub>3</sub>-33 (+0.11), and H<sub>2</sub>-34 (-0.18 and -0.09). Thus the absolute configurations at C-8 and C-24 and the relative stereochemistry of C-15, C-18, and C-19 of amphidinolide U were elucidated to be 1.

Amphidinolide U (1) is a novel 20-membered macrolide possessing a tetrahydrofuran ring, two *exo*-methylenes, three branched methyls, two ketones, two hydroxyl groups, and a  $C_{10}$  linear sidechain. The gross structure of C-9–C-29 unit of amphidinolide U (1) corresponds to that of C-14–C-34 of amphidinolide C (3), while the carbon skeleton of C-1–C-8 unit of 1 is very close to that of C-1 ~ C-8 of amphidinolide A<sup>9</sup> (4). This observation suggests that amphidinolide U (1) may be biogenetically related to amphidinolides C (3) and A (4). Amphidinolide U (1) exhibited weak cytotoxicity against murine lymphoma L1210 and human epidermoid carcinoma KB cells *in vitro* with IC<sub>50</sub> values of 12 and 20  $\mu$ g/mL, respectively.

## **Experimental Section**

**General Methods.** The IR and UV spectra were taken on a JASCO FT/IR-5300 and a JASCO Ubest-35 spectrophotometers, respectively. <sup>1</sup>H and 2D NMR spectra were recorded on a Bruker AMX-600 spectrometer, and <sup>13</sup>C NMR spectra were measured on a Bruker ARX-500 spectrometer. Positive-mode FAB mass spectra were obtained on a JEOL JMS HX-110 using *p*-nitrobenzyl alcohol as a matrix.

Cultivation and Isolation. The dinoflagellate *Amphidinium* sp. (strain number Y-56) was separated from the inside cells of the marine acoel flatworm *Amphiscolops* sp., which was collected off Zanpa, Okinawa. The dinoflagellate was unialgally cultured at 25 °C for two weeks in seawater medium enriched with 1% ES supplement. The harvested cells (420 g, wet weight, from 580 L of culture) were extracted with MeOH/toluene (3:1, 3 L x 3). After addition of 1 M NaCl aq. (1 L), the mixture was extracted with toluene (4 L x 3). The toluene-soluble fractions were evaporated under reduced pressure to give a residue (3.67 g), which was subjected to a silica gel column (CHCl<sub>3</sub>/MeOH, 98:2) and a Sep-Pak cartridge  $C_{18}$  (MeOH/H<sub>2</sub>O, 8:2) followed by  $C_{18}$  HPLC [LUNA C18(2), 5  $\mu$ m, Phenomenex®, 10 x 250 mm; eluent, CH<sub>3</sub>CN/H<sub>2</sub>O (75:25); flow rate, 2.5 mL/min; UV detection at 254 nm] to afford amphidinolides U (1, 0.8 mg, 0.0002 %, wet weight,  $t_R$  13.2 min), C (3, 0.0009 %,  $t_R$  16.5 min), and F (0.0006 %,  $t_R$  15.0 min).

**Amphidinolide** U (1): UV (MeOH)  $λ_{max}$  230 nm (ε 20000); IR (KBr)  $v_{max}$  3400, 2935, and 1710 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (Table 1); FABMS m/z 570 (M+H)<sup>+</sup> and 593 (M+Na)<sup>+</sup>; HRFABMS m/z 593.3662 [calcd for  $C_{34}H_{50}O_7Na$  (M+Na)<sup>+</sup>, 593.3645].

(S)-MTPA Ester (2a) of Amphidinolide U (1). To a CH<sub>2</sub>Cl<sub>2</sub> solution (10 µL) of amphidinolide U (1, 0.2 mg) were added 4-dimethylaminopyridine (0.01 mg), triethylamine (1 μL), and (R)-(-)-MTPACl (0.5 µL) at room temperature, and stirring was continued for 2 h. After addition of N,N-dimethyl-1,3-propanediamine (0.5  $\mu$ L) and evaporation of the solvent, the residue was passed through a silica gel column (hexane/EtOAc, 1:1) to afford the (S)-MTPA ester (2a, 0.1 mg) of 1. 2a: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.87 (3H, t, J = 7.3 Hz, H<sub>3</sub>-29), 1.09 (3H, d, J = 7.1 Hz, H<sub>3</sub>-32), 1.28 (2H, m, H<sub>2</sub>-28), 1.36 (2H, m, H<sub>3</sub>-27), 1.53 (1H, m, H-16b), 1.65 (3H, s, H<sub>3</sub>-31), 1.66 (1H, m, H-17b), 1.67 (3H, s,  $H_3$ -33), 1.83 (2H, m,  $H_2$ -26), 1.99 (1H, m, H-17a), 2.08 ~ 2.20 (5H, m,  $H_2$ -4, H<sub>2</sub>-5, and H-16a), 2.31 (1H, m, H-12b), 2.41 (1H, m, H-14b), 2.69 (1H, m, H-14a), 2.71 (1H, m, H-9b), 2.90 (1H, m, H-12a), 2.95 (1H, m, H-9a), 3.02 (1H, d, J = 16.3 Hz, H-2b), 3.08 (1H, d, J = 16.3 Hz, H-2b), 3. = 16.3 Hz, H-2a), 3.16 (1H, m, H-11), 3.53 (6H, s), 4.07 (m, H-18), 4.32 (1H, m, H-15), 4.91 (2H, s, H-30b and H-34b), 4.93 (2H, s, H-30a and H-34a), 5.21 (1H, m, H-19), 5.61 (1H, dd, J=1)7.9 and 15.1 Hz, H-20), 5.69 (1H, m, H-6), 5.78 (1H, s, H-24), 5.88 (1H, m, H-8), 6.11 (1H, d, J = 11.0 Hz, H-22), 6.50 (1H, dd, J = 11.0 and 15.1 Hz, H-21), 7.35 ~ 7.43 (6H, m), and 7.48 ~ 7.54 (4H, m); FABMS m/z 1025 (M+Na)<sup>+</sup>; HRFABMS m/z 1025.4220 [calcd for  $C_{54}H_{64}O_{11}F_7Na$  $(M+Na)^+$ , 1025.4250].

(*R*)-MTPA Ester (2b) of Amphidinolide U (1). Amphidinolide U (1, 0.2 mg) was treated with (*S*)-(+)-MTPACl (1  $\mu$ L) by the same procedure as described above to afford the (*R*)-MTPA ester (2b, 0.1 mg) of 1. 2b: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.88 (3H, t, J = 7.3 Hz, H<sub>3</sub>-29), 1.11 (3H, d, J = 7.1 Hz, H<sub>3</sub>-32), 1.28 (2H, m, H<sub>2</sub>-28), 1.42 (2H, m, H<sub>3</sub>-27), 1.49 (1H, m, H-16b), 1.56 (6H,

s, H<sub>3</sub>-31 and H<sub>3</sub>-33), 1.65 (1H, m, H-17b), 1.90 (2H, m, H<sub>2</sub>-26), 1.97 (1H, m, H-17a), 2.07 ~ 2.18 (5H, m, H<sub>2</sub>-4, H<sub>2</sub>-5, and H-16a), 2.36 (1H, m, H-12b), 2.42 (1H, m, H-14b), 2.67 (1H, m, H-14a), 2.81 (1H, m, H-9b), 2.95 (1H, m, H-12a), 3.01 (1H, m, H-9a), 3.03 (1H, d, J = 16.3 Hz, H-2b), 3.08 (1H, d, J = 16.3 Hz, H-2a), 3.13 (1H, m, H-11), 3.54 (6H, s), 4.05 (m, H-18), 4.32 (1H, m, H-15), 4.89 (1H, s, H-30b) 4.92 (1H, s, H-30a), 5.00 (1H, s, H-34b), 5.11 (1H, s, H-34a), 5.20 (1H, m, H-19), 5.50 (1H, dd, J = 7.9 and 15.1 Hz, H-20), 5.61 (1H, m, H-6), 5.76 (1H, s, H-24), 5.84 (1H, m, H-8), 5.98 (1H, d, J = 11.0 Hz, H-22), 6.44 (1H, dd, J = 11.0 and 15.1 Hz, H-21), 7.35 ~ 7.43 (6H, m), and 7.48 ~ 7.54 (4H, m); FABMS m/z 1025 (M+Na)\*; HRFABMS m/z 1025.4217 [calcd for  $C_{54}H_{64}O_{11}F_7Na$  (M+Na)\*, 1025.4250].

**Acknowledgment.** We thank Prof. T. Yamasu, University of the Ryukyus, and Dr. M. Ishibashi for help with dinoflagellate collection. This work was partly supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sports, and Culture of Japan.

## References

- 1 Ishibashi, M.; Kobayashi, J. Heterocycles 1997, 44, 543-572 and references cited therein.
- 2 Kobayashi, J.; Takahashi, M.; Ishibashi, M. Tetrahedron Lett. 1996, 37, 1449-1450.
- 3 Doi, Y.; Ishibashi, M.; Nakamichi, H.; Kosaka, T.; Ishikawa, T.; Kobayashi, J. J. Org. Chem. 1997, 62, 3820-3823.
- Kubota, T.; Tsuda, M.; Doi, Y.; Takahashi, A.; Nakamichi, H.; Ishibashi, M.; Fukushi, E.; Kawabata, J.; Kobayashi, J. *Tetrahedron*, 1998, 54, 14455-14464.
- Kobayashi, J.; Kubota, T.; Takahashi, M.; Ishibashi, M.; Tsuda, M.; Naoki, H. J. Org. Chem. 1999, 64, 1478-1482.
- 6 Kobayashi, J.; Ishibashi, M.; Wälchli, M. R.; Nakamura, H.; Hirata, Y.; Sasaki, T.; Ohizumi, Y. J. Am. Chem. Soc. 1988, 110, 490-494.
- 7 Kobayashi, J.; Tsuda, M.; Ishibashi, M.; Shigemori, H.; Yamasu, T.; Hirota, H.; Sasaki, T. J. Antibiot. 1991, 44, 1259-1261.
- 8 Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. J. Am. Chem. Soc. 1991, 113, 4092-4095
- Kobayashi, J.; Ishibashi, M.; Nakamura, H.; Ohizumi, Y.; Yamasu, T.; Sasaki, T.; Hirata, Y. Tetrahedron Lett. 1986, 27, 5755-5758.